



Comparison of test performance profile for blood tests of liver fibrosis in chronic hepatitis C

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Background/Aim: We evaluated the test performance profile (TPP) of blood tests of liver fibrosis.

Methods: Three hundred and fifty-six patients with C chronic hepatitis were included in two centers. Metavir staging of liver specimens by two independent pathologists and the following tests were evaluated: Fibrotest (FT), APRI, FibroMeter (FM), and Hepascore (HS).

Results: Metavir stages were: F0: 4%, F1: 55%, F2: 26%, F3: 11%, and F4: 4%. The AUROCs were not significantly different, respectively, FT, FM, APRI, HS: \geq F2: 0.79, 0.78, 0.76, 0.76; \geq F3: 0.81, 0.85, 0.81, 0.81; and F4: 0.86, 0.94, 0.92, 0.89. The TPP relies on the paired comparison of blood-test misclassification based on liver specimen, e.g. FT vs FM, respectively: F0+1: 18 vs 28% ($p = 0.0003$), \geq F2: 43 vs 31% ($p = 0.004$). There was no center effect.

Conclusions: In those populations, the four blood tests had a similar performance for significant fibrosis ($F \geq 2$), lying in the lower range of published results which is attributable to a low \geq F2 prevalence, and for \geq F3 and F4. However, FM and FT had performance profiles significantly different as a function of fibrosis stages or diagnostic target (fibrosis cut-off). This has to be considered during the interpretation process. Moreover, the performance should be reported with different diagnostic targets.

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Keywords: Liver fibrosis; Blood test; Metavir staging; Liver biopsy; C chronic hepatitis; Blood marker

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Abbreviations: APRI, aspartate aminotransferase to platelet ratio index; AUROC, area under the receiving–operating-characteristic; DA, diagnostic accuracy; FM, FibroMeter; FT, Fibrotest; HS, Hepascore; PACA, Provence-Côte d’Azur.

1. Introduction

Non-invasive diagnosis of liver fibrosis currently relies on blood tests and physical techniques like imaging and liver stiffness measurement. Several blood tests have been published [1]. The main diagnostic target criterion is clinically significant fibrosis. We can classify these blood tests into four categories: simple tests made

of a few indirect markers without algorithms like the aspartate aminotransferase to platelet ratio index (APRI) [2], indirect scores including indirect markers composite of an algorithm like Fibrotest (FT) [3], direct scores including direct markers – involved in connective tissue – composite of an algorithm like European liver fibrosis score [4] and mixed scores including indirect and direct markers composite of an algorithm like FibroMeter (FM) [5]. The performance of these tests largely depends on three main parameters: the accuracy of the reference method, usually liver biopsy, the intrinsic performance of the test and the prevalence of the diagnostic target. Although some characteristics of liver biopsy interpretation can be controlled (length, observer variability), this is the main limitation of performance measurement. The intrinsic performance mainly depends the blood markers and their measurement. Usually, such tests are implemented within one center with a training set and a validating set of patients. However, the next validating step is considered essential before expanding the tests to widespread routine use. Indeed, independent external validation includes assessing a potential variability of the three main parameters of diagnostic performance. Only, a few tests have been evaluated by external independent analysis with the exception of simple tests like APRI and an indirect test such as FT. However, recently some mixed tests with a high putative performance were published: Hepascore (HS) and FM [5,6]. In addition, the comparison of tests is often limited to simple descriptors whereas a more precise description of performance could be useful for clinical application [7].

The main aim of this study was to perform an external independent evaluation of these mixed tests in chronic hepatitis C by comparison with APRI as a simple test and FT as an indirect test. Direct tests were not evaluated since they include direct markers either with a low performance in chronic hepatitis C [4] or they are

not easily available in routine [8–10]. The secondary aim was to depict a new descriptor, the test performance profile.

2. Patients and methods

2.1. Patients

Three-hundred fifty-six patients attending the hepatogastroenterology unit of the University Hospital in Tours, and five units (two University Hospital, two public hospitals, one private clinic) from Provence-Côte d'Azur area, France, were retrospectively enrolled. Patients with chronic viral hepatitis C were included if they had a positive HCV-RNA in the serum and a liver biopsy and an alcohol consumption <30 g/day for the past 5 years. Blood fasting samples were taken at entry and a percutaneous (usually 1.4–1.6 mm diameter needle, suction technique) transcostal liver biopsy was performed within 1 week. Patients were not included if they had liver specimen <15 mm or other cause of liver disease or complicated cirrhosis or were given putative anti-fibrotic treatment (e.g. interferon or sartan) in the past 6 months. The study protocol was approved by a local Ethics committee and conformed to the ethical guidelines of the current Declaration of Helsinki. The dates of inclusion (liver biopsy) ranged from October 1994 to March 2004 in Tours center and from September 2002 to January 2004 in Provence area.

2.2. Methods

2.2.1. Blood parameters

Analyses of blood samples provided the following variables: platelet count, urea, bilirubin, γ -glutamyl transpeptidase, aspartate and alanine aminotransferases, prothrombin index, apolipoprotein A1, haptoglobin, hyaluronic acid, and α -2-macroglobulin. Blood markers were measured either on fresh blood or frozen sample of serum stored at -20°C . Techniques are detailed in Table 1. Sampling was performed for routine diagnostic aim within 1 week of liver biopsy.

2.2.2. Liver biopsy

Biopsy specimens were fixed in a formalin–alcohol–acetic solution and embedded in paraffin; 5 μm thick sections were stained with haematoxylin–eosin–safran. Fibrosis was staged by two independent expert pathologists according to the Metavir staging system [11]. Observers were blinded for patient characteristics. When the pathologists did not agree, the specimens were re-examined under a double-headed microscope to analyse discrepancies and reach a consensus.

Table 1

Analytical methods for blood tests: automates and reagents as a function of center

	Provence-Côte d'Azur	Tours
Aspartate aminotransferase	CX 7 (Beckman)	Hitachi/Olympus ^a
Alanine aminotransferase	CX 7 (Beckman)	Hitachi/Olympus
γ -Glutamyl transpeptidase	CX 7 (Beckman)	Hitachi/Olympus
Bilirubin	CX 7 (Beckman)	Hitachi/Olympus
Urea	CX 7 (Beckman)	CX 7 (Beckman)
Platelets	LH 750 (Coulter)	STKS (Beckman-Coulter)/LH750 (Beckman-Coulter) ^b
Prothrombin index	STAR	STA/STAR (neoplastine CI Diagnostica Stago) ^c
Apolipoprotein A1	BN Prospec (Dade Behring)	BN Prospec (Dade Behring)
Haptoglobin	BN Prospec (Dade Behring)	Array, Immage (Beckman-Coulter)
α -2-Macroglobulin ^d	BN Prospec (Dade Behring)	BN Prospec (Dade Behring)
Hyaluronate ^d	EIA Corgenix	EIA Corgenix

^a Hitachi until May 31, 2003.

^b SKTS until November 11, 2002.

^c STA until December 13, 2003.

^d Centralized dosage.

120 The pair of expert pathologists was different in each center including a
121 local pathologist and one out of two external pathologists from Beau-
122 jon Hospital for each center, i.e. a total of four pathologists. The rate
123 of initial observer disagreement was 38.8% in Provence area and 44.3%
124 in Tours ($p = 0.30$).

125 2.2.3. Statistical analysis

126 2.2.3.1. *Statistical tests.* Quantitative variables were expressed as
127 means \pm SD, unless otherwise specified. The prediction (or perfor-
128 mance) of each model is expressed either by the diagnostic accuracy
129 (DA), i.e. true positives and negatives, and by the area under the
130 receiving-operating-characteristic (AUROC). AUROCs were compared
131 by the Hanley-McNeil method for paired data [12]. Agreement
132 of qualitative variables was evaluated by the kappa index. Binary logistic
133 regression with forward stepwise inclusion of independent variables
134 was used. The cut-off of regression score was determined according to
135 the highest Youden index ($Se + Spe - 1$) to provide the best DA. The
136 statistical software used was SAS V8.2 (SAS Institute Inc., Cary, NC,
137 USA).

138 2.2.3.2. *Sample size calculation.* The size of population was
139 determined to show a significant difference between FM test and FT
140 [5]. With α risk: 0.05, β risk: 0.2, clinically significant fibrosis preva-
141 lence: 0.5, AUROC correlation: 0.70, and bilateral test, the sample size
142 was 360 patients for the following hypothesis of AUROC: FM: 0.85,
143 FT: 0.80.

144 3. Results

145 3.1. General characteristics

146 These characteristics are presented in Table 2. The
147 two populations were significantly different: the popula-
148 tion from Tours was younger with less severe hepatitis
149 and had a lower prevalence of significant fibrosis (31

vs 49%, $p = 0.001$) compared to the population from 150
Provence-Côte d'Azur. The date of liver biopsy was sig- 151
nificantly more recent in the Provence-Côte d'Azur 152
population. 153

3.2. Test performance 154

Indices are presented in Tables 3–5, respectively, for 155
significant fibrosis, severe fibrosis and cirrhosis. Signifi- 156
cant fibrosis includes F2 + F3 + F4, severe fibrosis 157
includes F3 + F4, and cirrhosis includes F4. The statisti- 158
cal comparison of performance between tests is classi- 159
cally only available with AUROC and is presented in 160
Table 6. There was no significant difference between 161
the four tests for the three diagnostic targets. 162

3.3. Pattern of test according to F stage 163

Fig. 1 depicts the box plots of blood tests for the 164
probability of significant fibrosis as a function of Meta- 165
vir fibrosis stage. It clearly shows a different pattern of 166
test probability. FT, FM and HS had a similar pattern 167
whereas APRI showed a narrower range for F0 and 168
F1. F2 patterns were similar for the four tests. F3 pat- 169
tern indicated a trend which was more evident in F4 170
stage: tests including hyaluronic acid (FM and HS) dis- 171
played a higher probability value and a narrower range 172
than others which was especially evident for FM in F4. 173
These observed differences lead us to compare the per- 174
formance within each Metavir F stage. This was made 175

Table 2
General characteristics of patient populations

	Whole population	Provence-Côte d'Azur	Tours	<i>p</i>
No. of patients	356	198	158	–
Sex (% male)	189 (53)	99 (50)	90 (57)	0.23
Age (year)	44.9 \pm 12.9	48.1 \pm 13.7	40.9 \pm 10.5	0.01
Metavir fibrosis stage				
F0 (%)	15 (4)	9 (4)	6 (4)	0.79
F1 (%)	195 (55)	92 (48)	103 (65)	0.002
F2 (%)	95 (26)	61 (31)	34 (22)	0.08
F3 (%)	38 (11)	25 (12)	13 (8)	0.29
F4 (%)	13 (4)	11 (6)	2 (1)	0.07
Clinically significant fibrosis (%)	146 (41)	97 (49)	49 (31)	0.001
Severe fibrosis (%)	51 (14)	36 (18)	15 (9.5)	0.02
Metavir fibrosis score	1.5 \pm 0.9	1.7 \pm 0.9	1.4 \pm 0.7	0.001
Liver specimen length (mm)	22.0 \pm 7.1	23.3 \pm 7.3	20.5 \pm 6.4	0.04
Median liver biopsy date	15 January 2003	20 May 2003	8 April 1999	<.0001
Platelets (G/L)	211.2 \pm 65.0	203.2 \pm 60.3	221.2 \pm 69.2	0.64
Prothrombin index (%)	92.8 \pm 7.6	92.1 \pm 7.7	93.7 \pm 7.4	0.03
ASAT (UI/L)	49.3 \pm 37.3	57.3 \pm 42.7	39.2 \pm 26.0	<.0001
ALAT (UI/L)	76.5 \pm 66.2	77.7 \pm 71.3	74.9 \pm 59.3	0.99
GGT (UI/L)	67.1 \pm 108.7	75.0 \pm 117.3	57.3 \pm 96.5	0.10
Bilirubin (μ mol/L)	10.5 \pm 5.3	9.9 \pm 5.5	11.0 \pm 5.0	0.03
Urea (mmol/L)	5.8 \pm 1.6	6.0 \pm 1.8	5.6 \pm 1.3	0.11
Apolipoprotein A1 (g/L)	1.5 \pm 0.4	1.5 \pm 0.4	1.4 \pm 0.4	0.05
Haptoglobin (g/L)	0.94 \pm 0.45	0.93 \pm 0.45	0.96 \pm 0.46	0.59
α -2-Macroglobulin (mg/dL)	287 \pm 111	272 \pm 91	305 \pm 130	0.10
Hyaluronate (μ g/L)	52.4 \pm 96.2	64.9 \pm 114.7	36.7 \pm 63.2	<.0001

Table 3
Performance of blood tests for significant fibrosis

Test	Cut-off	Se	Spe	Positive PV	Negative PV	Positive LR	Negative LR	DA	AUROC (95% CI)
FibroMeter	0.57	64	81	70	77	3.30	0.44	74	0.78 (0.73;0.82)
Fibrotest	0.44	67	80	70	78	3.36	0.41	74	0.79 (0.75;0.83)
Hepascore	0.32	77	63	59	80	2.09	0.37	69	0.76 (0.71;0.80)
APRI	0.39	77	66	61	80	2.24	0.35	70	0.76 (0.72;0.81)

Se, sensitivity; Spe, specificity; PV, predictive value; LR, likelihood ratio; DA, diagnostic accuracy; AUROC, area under the receiving–operating–characteristic.

Table 4
Performance of blood tests for severe fibrosis

Test	Cut-off	Se	Spe	Positive PV	Negative PV	Positive LR	Negative LR	DA	AUROC (95% CI)
FibroMeter	0.67	82	76	37	96	3.44	0.23	77	0.84 (0.80;0.88)
Fibrotest	0.45	84	69	31	96	2.74	0.23	71	0.81 (0.77;0.85)
Hepascore	0.53	78	72	32	95	2.78	0.30	73	0.81 (0.76;0.85)
APRI	0.58	75	76	34	95	3.11	0.34	76	0.81 (0.76;0.85)

Se, sensitivity; Spe, specificity; PV, predictive value; LR, likelihood ratio; DA, diagnostic accuracy; AUROC, area under the receiving–operating–characteristic.

Table 5
Performance of blood tests for cirrhosis

Test	Cut-off	Se	Spe	Positive PV	Negative PV	Positive LR	Negative LR	DA	AUROC (95% CI)
FibroMeter	0.88	92	87	21	100	7.20	0.09	87	0.94 (0.91;0.96)
Fibrotest	0.56	85	74	11	99	3.19	0.21	74	0.86 (0.82;0.89)
Hepascore	0.61	92	72	11	100	3.30	0.11	73	0.89 (0.86;0.92)
APRI	0.83	100	83	18	100	5.81	0.00	83	0.92 (0.88;0.94)

Se, sensitivity; Spe, specificity; PV, predictive value; LR, likelihood ratio; DA, diagnostic accuracy; AUROC, area under the receiving–operating–characteristic.

Table 6
Comparison of AUROC between blood tests (*p* value of Hanley–McNeil test)

	Significant fibrosis (\geq F2)	Severe fibrosis (\geq F3)	Cirrhosis (F4)
FM vs FT	0.67	0.32	0.09
FM vs HS	0.28	0.19	0.19
FM vs APRI	0.46	0.23	0.63
FT vs APRI	0.35	0.89	0.40
FT vs HS	0.08	0.91	0.40
HS vs APRI	0.84	0.95	0.73

FM, FibroMeter; FT, Fibrotest; HS, Hepascore.

176 possible by first calculating the disagreement rate
 177 between each blood test and liver specimen for the diag-
 178 nosis of significant fibrosis. Then, the disagreement rates
 179 were compared between a pair of blood tests by the
 180 McNemar test for paired comparison. We focused the
 181 presentation of test performance profile to the two tests
 182 with the nearer and highest AUROCs, i.e. FM and FT.
 183 Table 7 shows a fair agreement between both tests.
 184 More importantly, it clearly shows a significantly differ-
 185 ent test performance profile. FT was superior to FM for

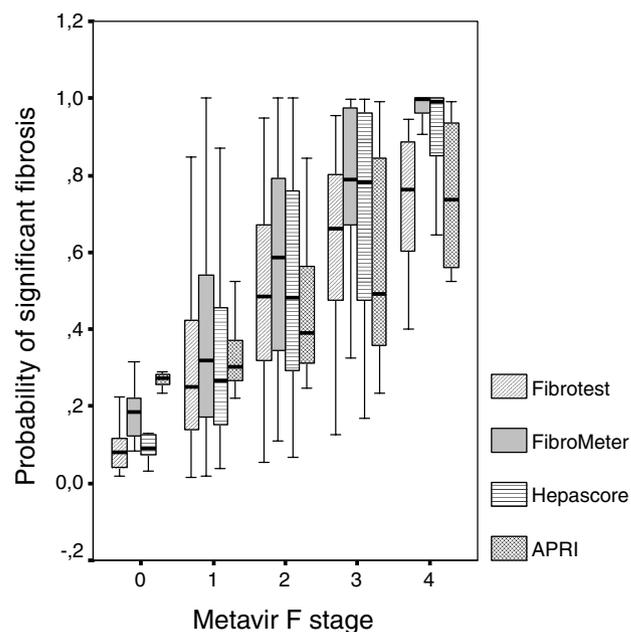


Fig. 1. Box plots (median, quartiles and extremes) of blood tests as a function of Metavir fibrosis stage for probability of significant fibrosis. APRI was normalized using logistic regression.

Table 7
Rate (%) of misclassified patients for significant fibrosis according to Metavir fibrosis stage for FT and FM

	Metavir fibrosis stage								
	0	1	2	3	4	<2	≥2	≥3	All
No. of patients	15	195	95	38	13	210	146	51	356
Fibrotest	0	19.0	51.6	28.9	15.4	17.6	42.5	25.5	27.8
FibroMeter	0	29.7	41.1	15.8	0	27.6	30.8	11.8	28.9
<i>p</i> (McNemar)	— ^a	0.0003	0.03	0.18	— ^a	0.0003	0.004	0.065	0.734
Kappa	— ^a	0.383	0.539	0.335	— ^a	0.396	0.520	0.310	0.461

^a Calculation impossible.

186 F1 stage whereas FM was superior to FT for F2 or F3
 187 or F4 stages. Finally, FT was significantly lower than
 188 FM for the misclassification rate in patients without sig-
 189 nificant fibrosis (18 vs 28%, respectively, $p = 0.0003$)
 190 whereas FM was significantly lower than FT for the mis-
 191 classification rate in patients with significant fibrosis (31
 192 vs 42.5%, respectively, $p = 0.004$). Fig. 2 clearly shows
 193 the different test performance profiles between FT and
 194 FM.

195 3.4. Center effect

196 The differences of test performances according to
 197 each center are shown for AUROC in Table 8. Values
 198 were similar except for F4 but only 2 patients had cir-
 199 rhosis in one center which precluded a reliable compar-
 200 ison. It should be stated that there is no statistical test to
 201 compare unpaired AUROC. Therefore, the center effect
 202 was tested using forward logistic regression by including
 203 the variables significantly different between centers as
 204 independent variables and disagreement for significant
 205 fibrosis between blood tests and liver specimen as depen-
 206 dent variable. For FM, the only independent variable
 207 was age ($p = 0.002$) whereas center had no significant

effect ($p = 0.90$). For FT, the only independent variable
 208 was Metavir F stage ($p = 0.001$) whereas center had no
 209 significant effect ($p = 0.43$) with a borderline effect for
 210 age ($p = 0.058$) and sex ($p = 0.088$). So, the center had
 211 no independent effect on test performance, the differenc-
 212 es between centers being due to demographic differences,
 213 age or fibrosis stage, both being significantly related
 214 ($p < 10^{-4}$ by ANOVA).
 215

216 3.5. Effect of liver specimen size

217 There was no correlation between length of liver spec-
 218 imen and Metavir F stage ($r = -0.065$, $p = 0.22$). AUR-
 219 OCs were evaluated as a function of the liver specimen
 220 median (21.0 mm) in Table 9. AUROCs were not
 221 improved in the largest specimens.

222 4. Discussion

223 4.1. Comparison with previous studies

224 APRI and FT have been evaluated in several inde-
 225 pendent studies [5,13]. However, truly independent stud-
 226 ies mainly devoted to validation are rare [14,15]. In the
 227 present study, a lower performance was observed for
 228 the 4 blood tests (AUROC for significant fibrosis:
 229 0.76–0.79) than in the original publications where AUR-
 230 OCs were 0.80–0.88 (test-validation) for APRI [2], 0.85
 231 for HS [6], 0.84–0.87 for FT [3] and 0.88–0.91 for FM
 232 [5]. But our results fit well with the 0.77 median
 233 AUROC reported for 10 different blood tests in valida-
 234 tion populations vs 0.81 in training populations [7]. So,
 235 the expected difference between FT and FM [5] was not
 236 observed. The main cause was a less severe fibrosis stage
 237 and a younger age. This was observed in one center
 238 compared to the other. In addition, the prevalence of
 239 significant fibrosis in both centers was low: 41% here
 240 vs 56% in the original FM study [5] and 49% in the ini-
 241 tial FT study [3], 44–57% in the original HS study [6]
 242 and 47–50% in the original APRI study [2]. The interac-
 243 tion between this prevalence and AUROC, depicted in
 244 the present study, shows that the sample size calculation
 245 is hindered by this confusing factor.

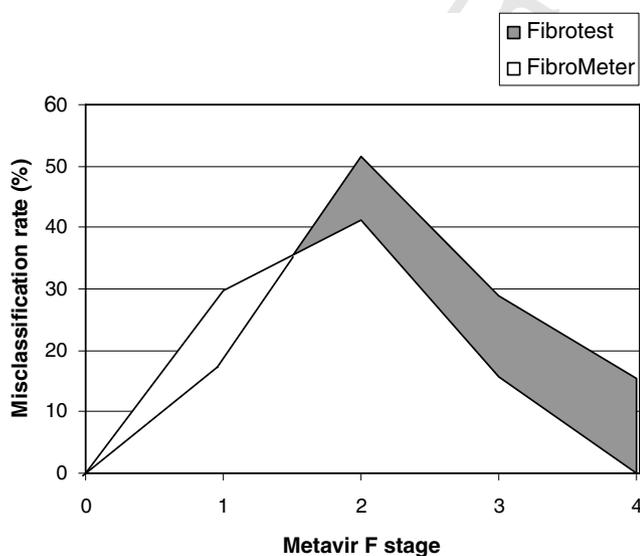


Fig. 2. Profile of test performance: comparison of misclassification rate for significant fibrosis between FT and FM.

Table 8
Comparison of AUROC for significant fibrosis between the two centers

Center	Significant fibrosis (\geq F2)		Severe fibrosis (\geq F3)		Cirrhosis (F4)	
	PACA	Tours	PACA	Tours	PACA	Tours
No. of patients with diagnostic target	97	49	36	15	11	2
FibroMeter	0.767	0.773	0.852	0.800	0.911	1
Fibrotest	0.805	0.789	0.815	0.817	0.844	0.920
Hepascore	0.774	0.758	0.819	0.798	0.893	1
APRI	0.762	0.736	0.804	0.772	0.872	0.926

No statistical comparison available for unpaired data.
PACA, Provence-Côte d'Azur.

246 In the present study, we have implemented a new
247 method to compare the test performance. The test per-
248 formance profile relies on the comparison of misclassifi-
249 cation rates, based on liver specimens, that are tested by
250 a paired McNemar test. This method, only applied here
251 to FM and FT, clearly shows that the test performance
252 depends on the fibrosis stage (Table 7). Thus, both FT
253 and FM displayed a peak of misclassification rate for
254 F2 (Fig. 2). Moreover, the test performance profile
255 was different since this peak was significantly lower for
256 FM and the values were significantly lower with FM
257 at the right of this peak, i.e. for all stages of clinically
258 significant fibrosis. This explains that the difference in
259 test performance between different blood tests depends
260 on the relative prevalence of fibrosis stages (e.g., a pop-
261 ulation with a high prevalence of clinically significant
262 fibrosis or severe fibrosis will favour FM compared to
263 FT and *vice versa*).

264 Finally, the box plots of blood tests against fibrosis
265 stage clearly show the diagnostic robustness of a test
266 for a fibrosis stage, e.g. the range of FM for F4 is very
267 narrow (Fig. 1) that suggests the robustness of this test
268 to correctly classify F4 stage in different populations.
269 In addition, the comparison of box plots shows that
270 the score probabilities of different tests have not a simi-
271 lar prediction for the fibrosis stage, e.g. the median value
272 of FM for F3 is slightly superior to that of FT for F4.
273 Thus, the numerical values of different tests are not
274 comparable.

275 4.2. Factors influencing the test performance

276 We have previously shown that the test performance
277 depends on the fibrosis stage. In addition, the perfor-

278 mance of tests globally depends on the diagnostic target
279 as determined by Metavir F stage cut-off (Tables 3–5):
280 the greater the diagnostic target, the higher the observed
281 performance. Thus, on one hand, the absolute test per-
282 formance increased as a function of the diagnostic tar-
283 get. On the other hand, despite a low prevalence of
284 significant fibrosis in the present study, the relative
285 FM performance, compared to that of FT, increased
286 as a function of the diagnostic target. These observa-
287 tions depict, with different shades, a test performance
288 profile as clearly observed in Fig. 2. This profile should
289 be adjusted. Indeed, this profile is based on a liver spec-
290 imen as reference. However, the reliability of such a ref-
291 erence, as shown by the interobserver disagreement,
292 displays the same V pattern [16] suggesting that a signif-
293 icant part of the misclassification rate is attributable to
294 misclassification by liver biopsy interpretation itself.
295 Nevertheless, this profile is correct in interpreting the
296 differences between blood tests. Thus, both FM and
297 FT are characterized by a 100% accuracy in F0 whereas
298 this figure is attained in F4 only with FM.

299 This new descriptor of test performance can be
300 applied to compare different populations. Thus, the
301 prevalence of fibrosis stage depends on the cause of
302 chronic liver disease, e.g. in co-infections with HIV,
303 the prevalence of F0 is usually low and that of F4 is high
304 whereas the test performance might depend on specific
305 variation of blood markers in HIV population [17,18]
306 thus confusing the comparison of global test perfor-
307 mance between mono-infected and co-infected popula-
308 tions. The use of the test profile contributes to clarify
309 the comparison by deleting this confounding factor
310 and making possible a direct comparison of test perfor-
311 mance for each fibrosis stage.

Table 9
AUROC as a function of the liver specimen median according to diagnostic targets

Liver specimen length (mm)	Significant fibrosis		Severe fibrosis		Cirrhosis	
	<21	\geq 21	<21	\geq 21	<21	\geq 21
No. of patients with diagnostic target	67	79	29	22	10	3
FibroMeter	0.798	0.757	0.854	0.830	0.952	0.942
Fibrotest	0.799	0.787	0.851	0.765	0.893	0.750
Hepascore	0.770	0.746	0.828	0.788	0.910	0.864
APRI	0.753	0.775	0.826	0.787	0.940	0.875

312 This study also shows that there was no center effect
 313 on the AUROC of the four blood tests and no signifi-
 314 cant center effect on the misclassification rate. In fact,
 315 this misclassification rate was independently influenced
 316 by patient age or the fibrosis stage which were signifi-
 317 cantly different as a function of centers. This clearly sug-
 318 gests that the difference between centers was not due to
 319 difference in analytical characteristics of blood determi-
 320 nations. The lower prevalence of significant fibrosis in
 321 the two centers compared to previous studies [2–5] can
 322 be attributed either to the centers themselves, e.g. the
 323 Provence-Côte d’Azur area included units closer to the
 324 general population of chronic HCV patients cared for
 325 by hepato-gastroenterologists. It can be also attributed
 326 to the more recent inclusion period compared to original
 327 studies since the degree of fibrosis stage decreases as a
 328 function of dates [19].

329 The blood test accuracy is also limited by the sampling
 330 error of liver biopsy. This could be partially circumvented
 331 by using only the largest liver specimens. However, in the
 332 present study, there was no correlation between length of
 333 liver specimen and Metavir F stage unlike in other studies
 334 [16,20]. Moreover, AUROCs were not improved in larg-
 335 est specimens (Table 8). Finally, recent studies have sug-
 336 gested that blood markers of liver fibrosis combined
 337 either in a stepwise algorithm [21] or to elastometry [22]
 338 might improve the diagnostic performance.

339 4.3. What should be the ideal test profile?

340 Of course an ideal test would have a 100% diagnos-
 341 tic accuracy. However, this will be very difficult to
 342 achieve as long as liver biopsy remains the reference.
 343 Meanwhile, if we have to choose a blood score among
 344 others, the test performance profile described here con-
 345 tributes to facilitate that choice. Indeed, the test profile
 346 suggests that a global performance like diagnostic
 347 accuracy or AUROC is not sufficient for comparison
 348 since it is too dependent on the prevalence of fibrosis
 349 stage. The inverse significant differences in misclassifi-
 350 cation rate between FT and FM for the low or the
 351 high-fibrosis stages validate the use of a test perfor-
 352 mance profile depicting the accuracy (or its inverse:
 353 the misclassification rate) as a function of fibrosis
 354 stage. In the present case, we have the choice between
 355 a test overestimating low stages and better classifying
 356 high stages of fibrosis and a test better classifying the
 357 early stage and underestimating high stages of fibrosis.
 358 We consider that the first profile is convenient for a
 359 diagnostic goal, since it is important to correctly classi-
 360 fy severe fibrosis or cirrhosis for complication screen-
 361 ing, as well in a screening perspective since
 362 overestimation means higher sensitivity which is suit-
 363 able for a fibrosis screening. However, this last goal
 364 has to be validated in an adequate population. Recent-
 365 ly, improved test reporting was recommended for the

description of blood tests including likelihood ratios 366
 and diagnostic odds ratio [7]. We suggest that this test 367
 performance profile should be added to this descrip- 368
 tion. Indeed, test performance profile allows two new 369
 kinds of description of diagnostic accuracy for a given 370
 diagnostic target and a test based on a semiquantitative 371
 classification: first, detailed accuracy as a function of 372
 stages for a single test; second, comparison between 373
 two tests using diagnostic accuracy either of all the 374
 stages (global test) and/or stage by stage of fibrosis. 375
 We suggest that test performance profile should be 376
 used in addition to AUROC, positive and negative pre- 377
 dictive values, especially as main descriptor in statisti- 378
 cal comparisons besides AUROC. But both are 379
 limited by paired comparison, i.e. restricted to the same 380
 sample. 381

382 In conclusion, this study shows a good performance
 383 of the four blood tests but lower than in the original
 384 publications. We have used a new method to compare
 385 the diagnostic accuracy as a function of fibrosis stage.
 386 This pattern is called test performance profile and shows
 387 significant differences in diagnostic accuracy between
 388 blood tests as a function of fibrosis stage. This test per-
 389 formance profile should be added in the description of
 390 test performance especially in comparative studies.
 391 Finally, the test performance should be reported for dif-
 392 ferent diagnostic targets.

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